

# Advances in eDNA monitoring of marine mammals: Current capabilities and future directions

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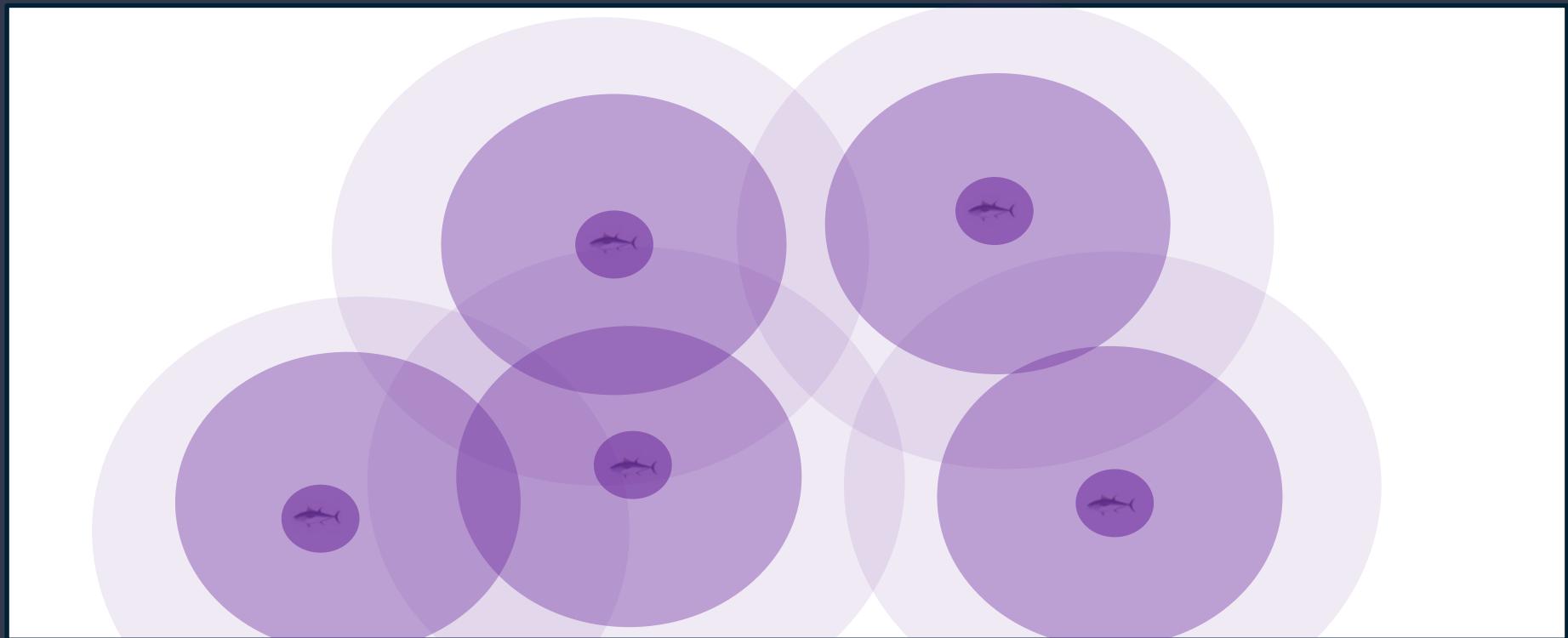
eDNA = genetic traces recovered from the water

But what does it mean? What are we sampling?



More of a species = more of its DNA (in the world)

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Organisms are discrete; eDNA is more continuous

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# Forms of Information from eDNA (+ eRNA)

Detection

Concentration

eDNA (various fragments)

eRNA (various fragments)

Sequence Information

Haplotypes

Functional genetics + expression

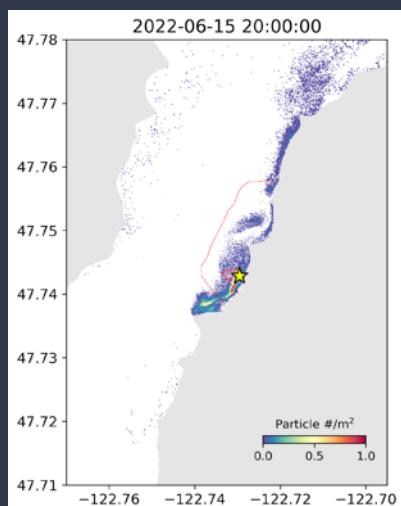
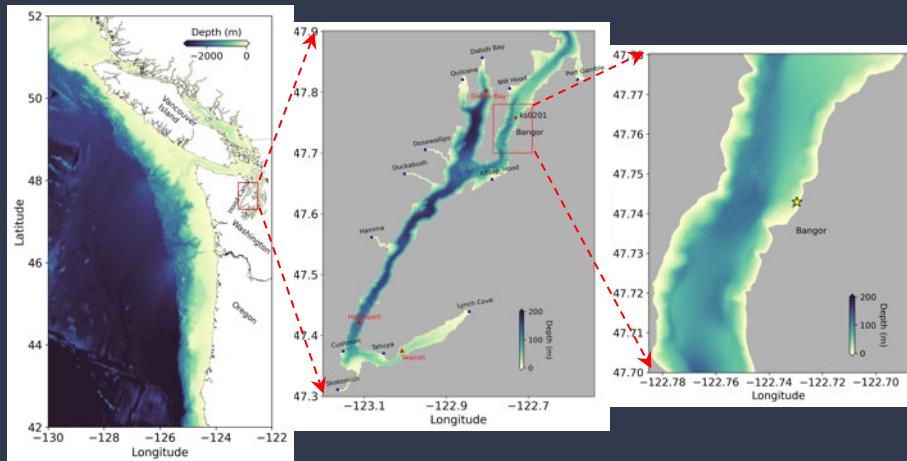
Methylation

Challenge: Linking this information to parameters of interest in the real world

# Cetacean eDNA in Space and Time

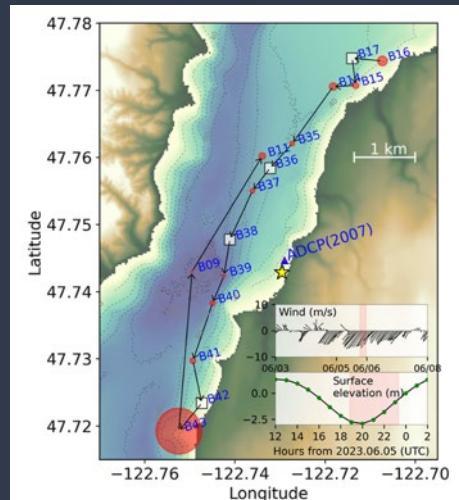
Captive *Tursiops* as model

High-res  
oceanographic  
model

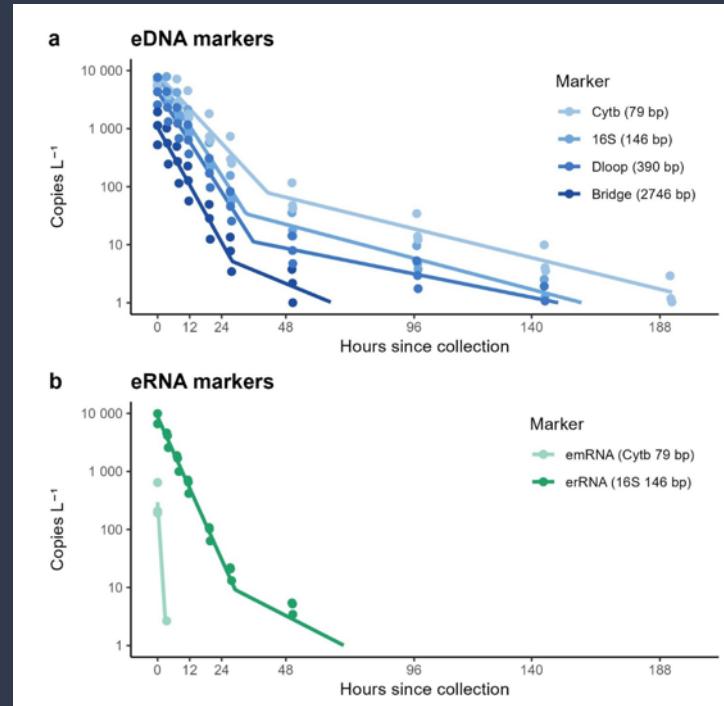
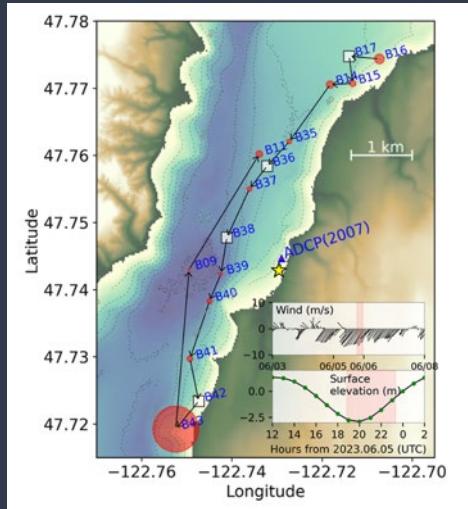


Particle tracks predict  
eDNA dispersal

Expected vs real-world observations



# Cetacean eDNA in Space and Time



Brendão et al. 2025

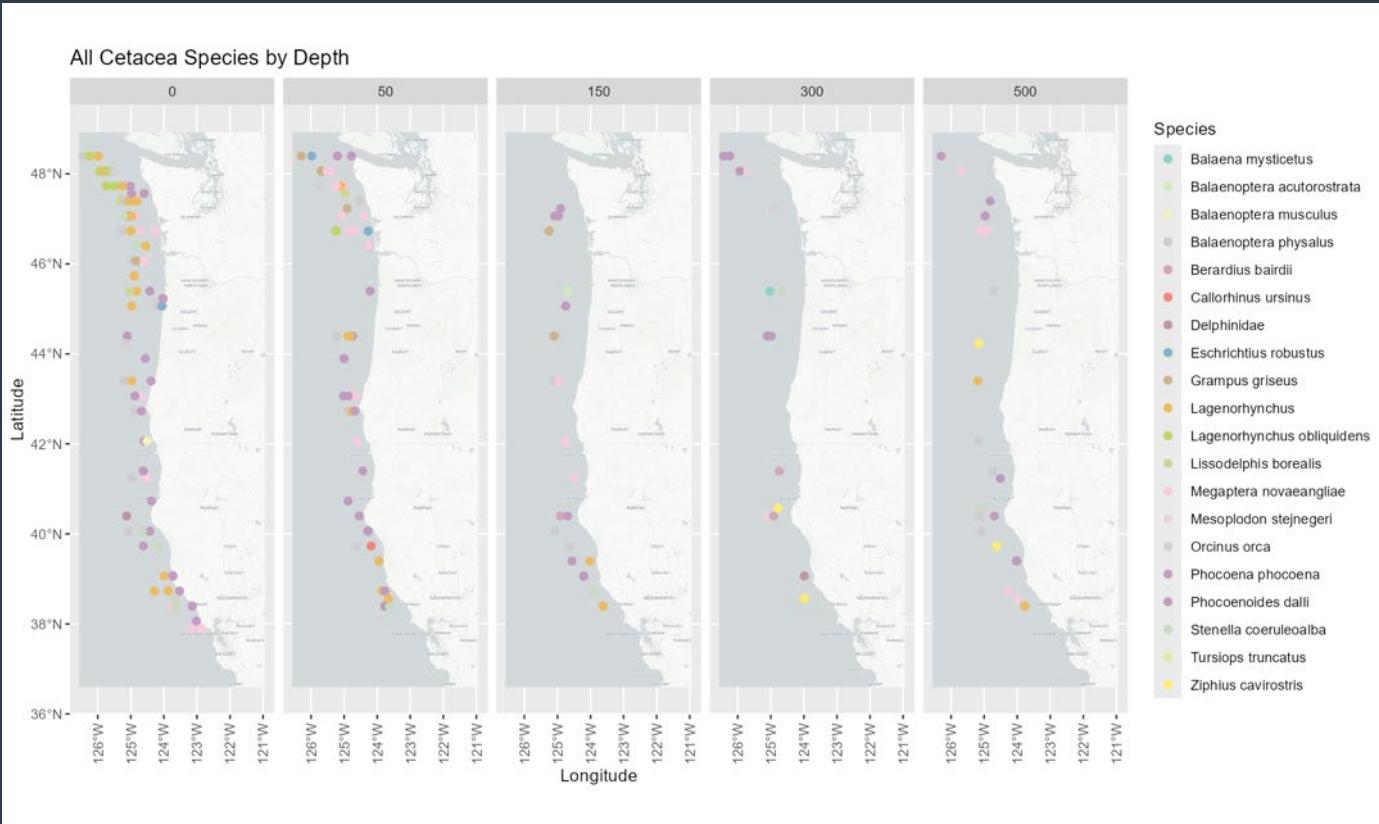
At scales of 1-10km:

- we can trace eDNA dispersal via oceanography
- eRNA degrades very quickly; eDNA pretty quickly

# Cetacean eDNA at Large Spatial Scales

West coast samples via NOAA

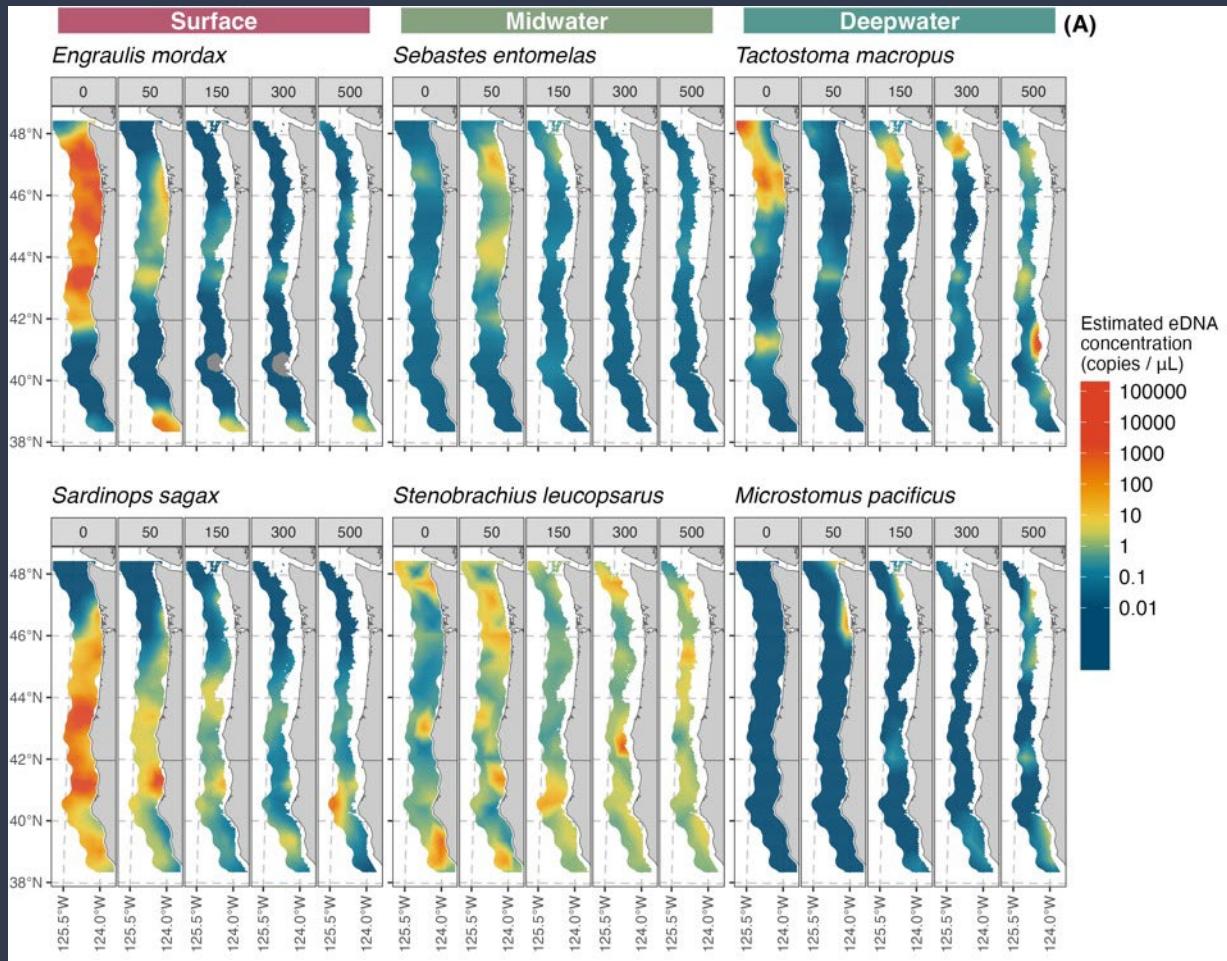
50% of all samples have cetacean sequences



# Next Up

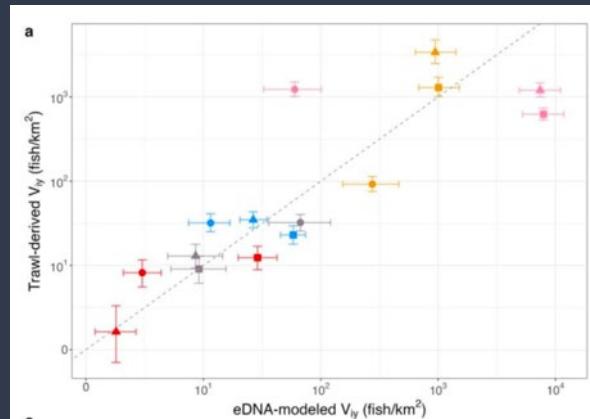
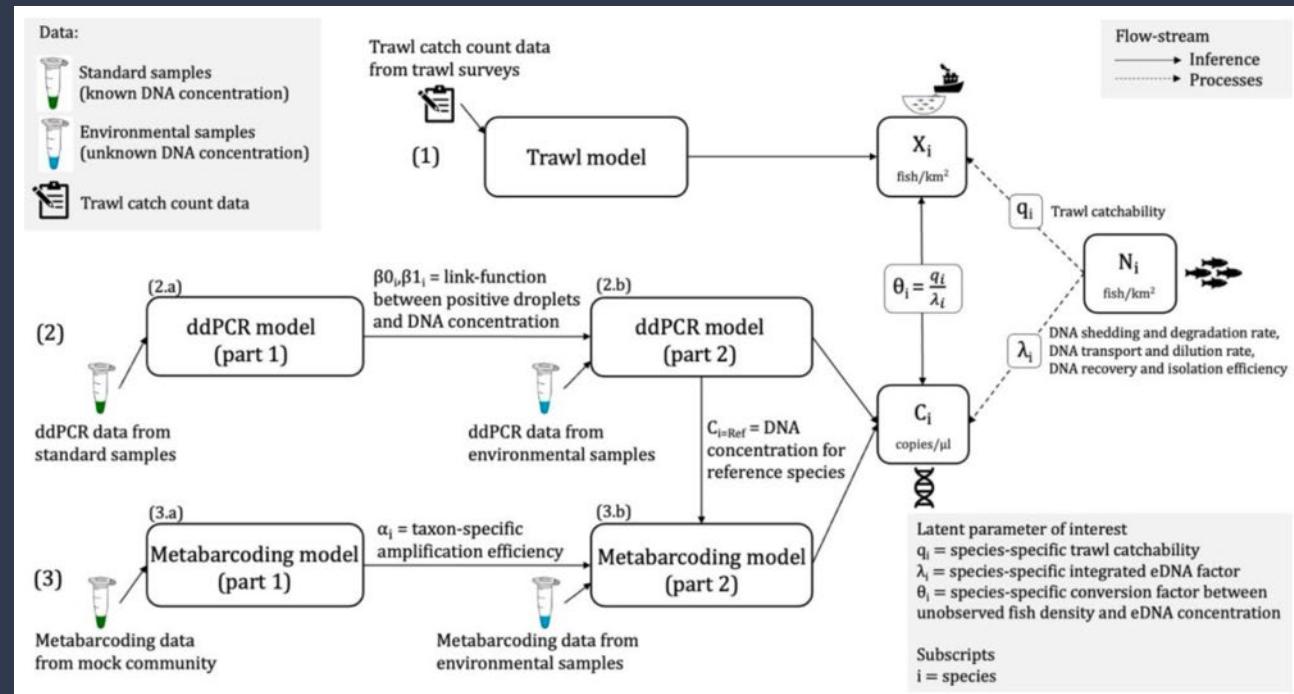
Quantifying and Smoothing

Applying fisheries work to  
cetaceans



# Next Up

Jointly estimating cetacean abundance  
from visual, acoustic, and eDNA  
observations



Reciprocal out-of-sample cross-validation

# Thanks!

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Ole Shelton

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Shana Hirsch

Aden Ip

Owen Liu

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Krista Nichols

Olivia Scott

Stephanie Matthews

Len Thomas

Brice Semmens

Simone Baumann-Pickering

Michaela Alksne

Nastassia Patin

The David and Lucile Packard Foundation

Oceankind

Office of Naval Research

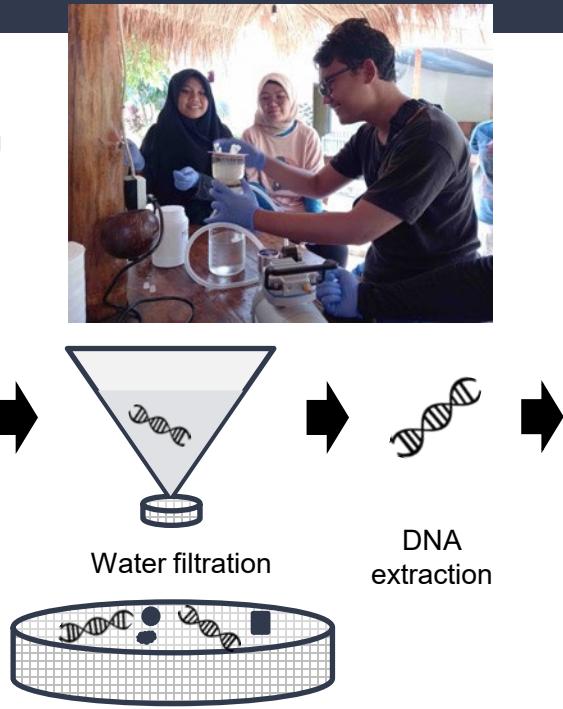
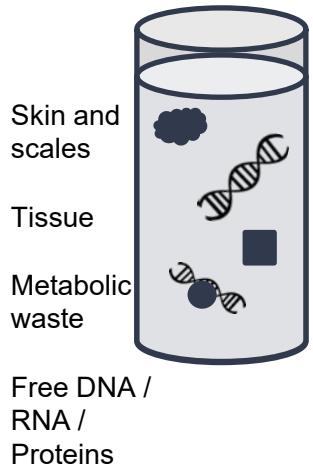
NOAA Fisheries

# Extra Slides

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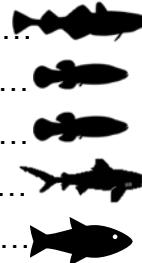
# What is eDNA?

A soup of information



Metabarcoding:  
Hundreds/thousands of species,  
less quantitative



GAATCGC...  
GTCGATA...  
GTCGATA...  
CTCGATA...  
AGCACTC...  


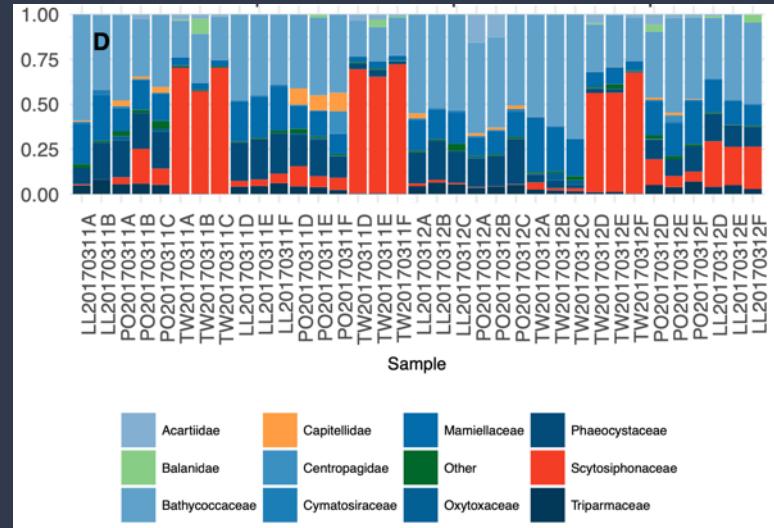
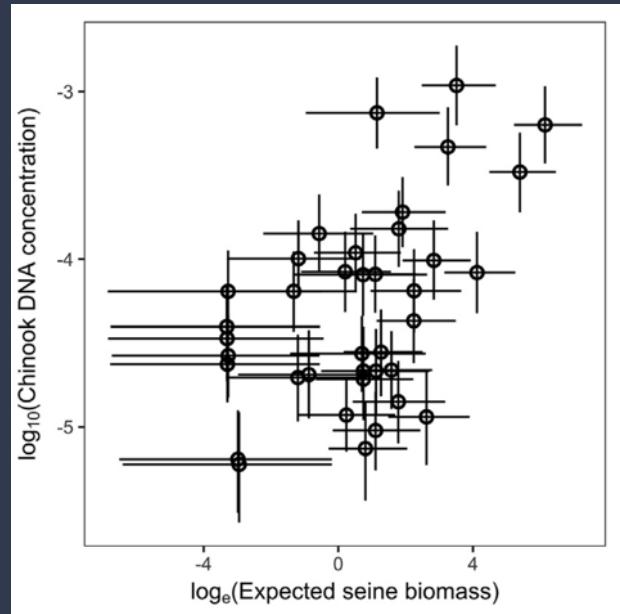
Scalable  
Automatable  
Sensitive  
Non-invasive

Quantitative PCR:  
single-target, quantitative



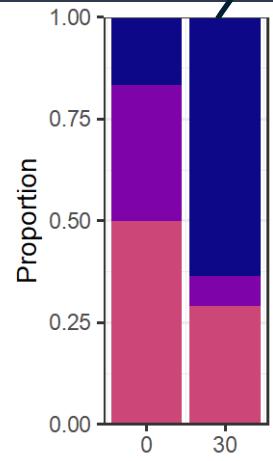
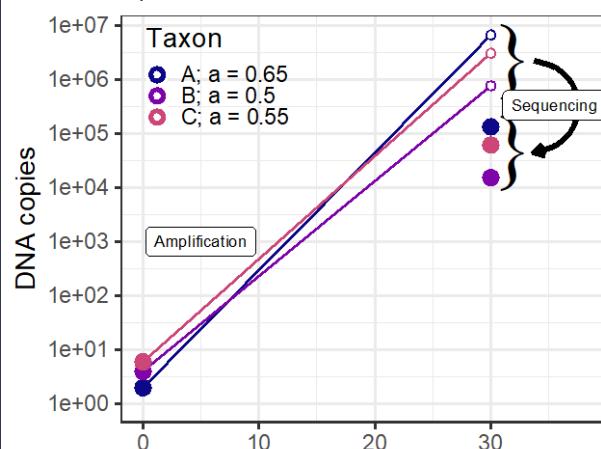
AGCACTC...  


# Single-Species (qPCR / ddPCR) VS. Multi-Species (Metabarcoding)



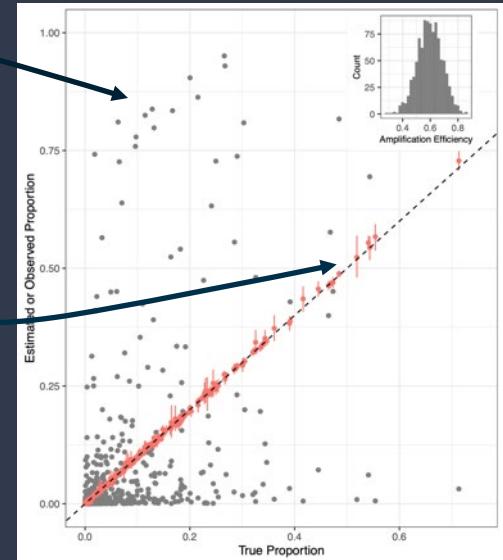
# Important Note on Metabarcoding

Amplification Bias: Different species may/do amplify at different rates



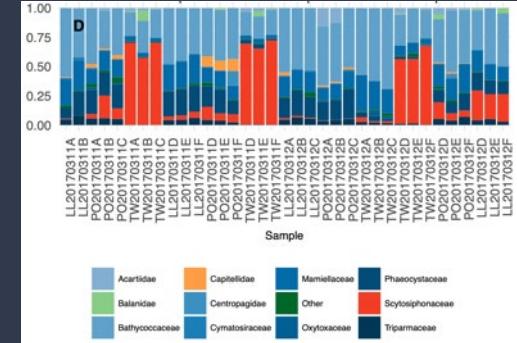
Uncalibrated

Calibrated



# Other Important Note on Metabarcoding

The resulting dataset is compositional and contains little/no info about absolute concentration



$$\mathbf{Y} \sim \text{Multinomial}(\boldsymbol{\mu}, N),$$

$$\mu_i = \frac{e^{\nu_i}}{\sum_{i=1}^I e^{\nu_i}},$$

$$\nu_i = \beta_i + N_{\text{PCR}} \alpha_i + \epsilon_i,$$

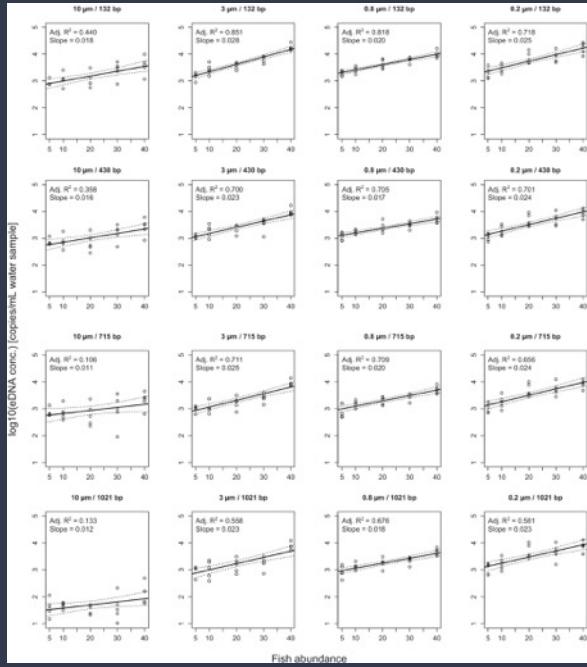
$$\epsilon_i \sim N(0, \tau_i),$$

- We model the PCR process for each species in a composition
- Getting to absolute abundance requires additional information
  - e.g., visual counts, trawl, trap, whatever

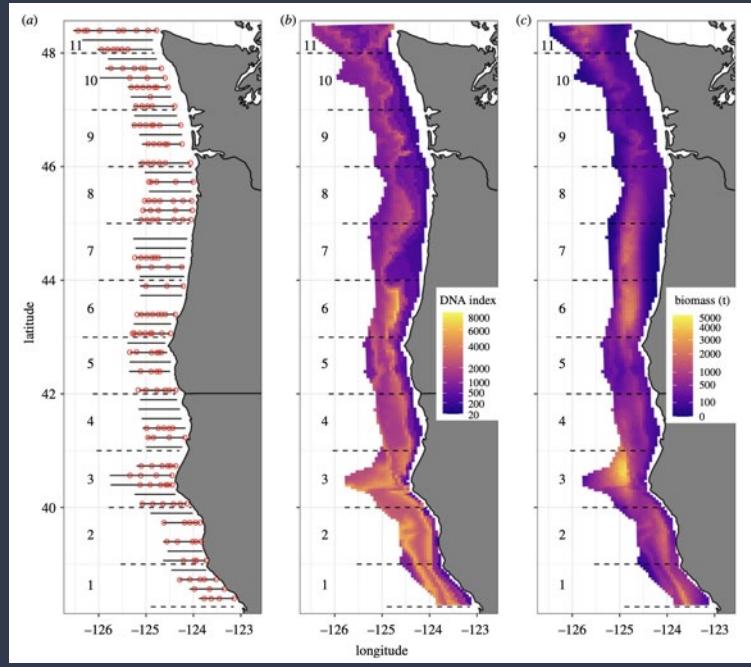
# Single Species – qPCR / ddPCR

Quantitative estimates of eDNA concentration

Tiny Scale



Large Scale



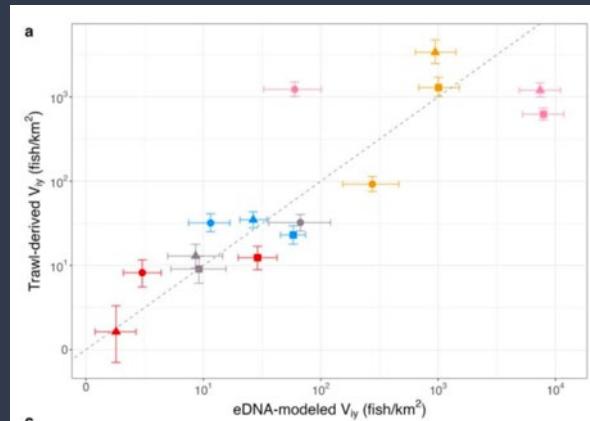
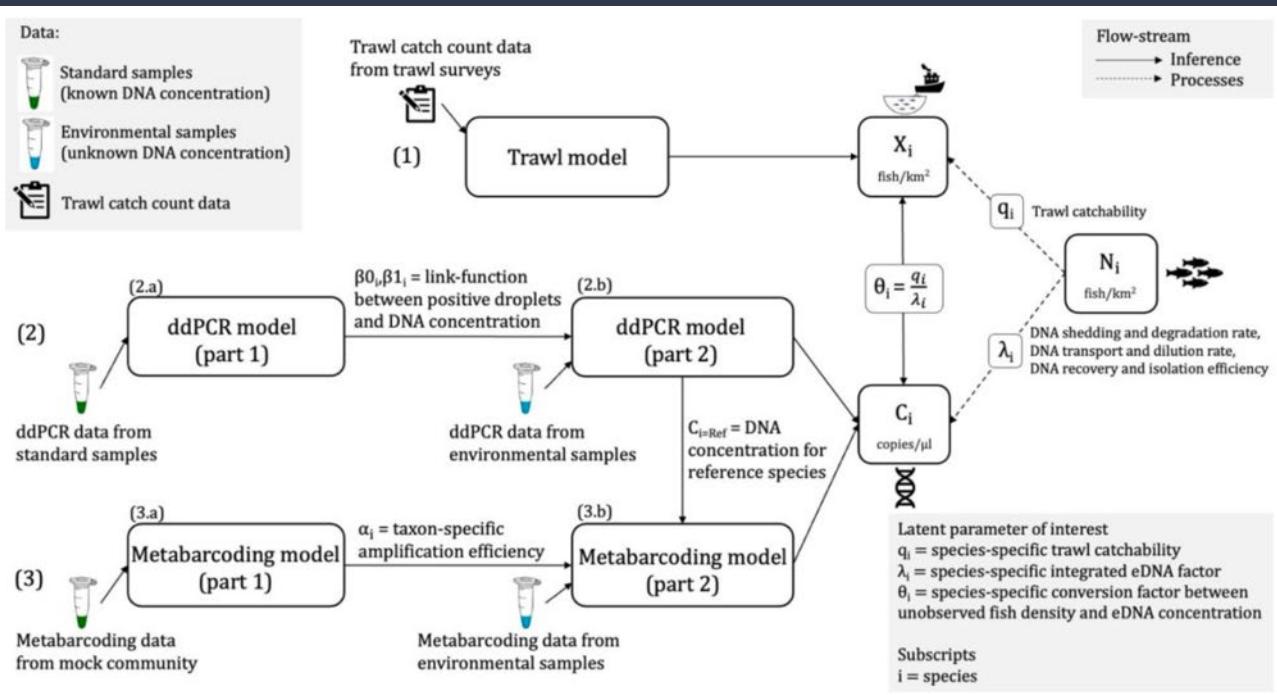
Jo and Yamanaka (2022) – Zebrafish aquarium study across pore sizes + fragment sizes.

Shelton et al. (2022): smoothed qPCR signal reflects Hake (Merluccius productus) eDNA distribution

# Multispecies (Metabarcoding)

Metabarcoding (proportions) + dPCR (ref species quantity) + trawls

Quantitative estimates of species abundance



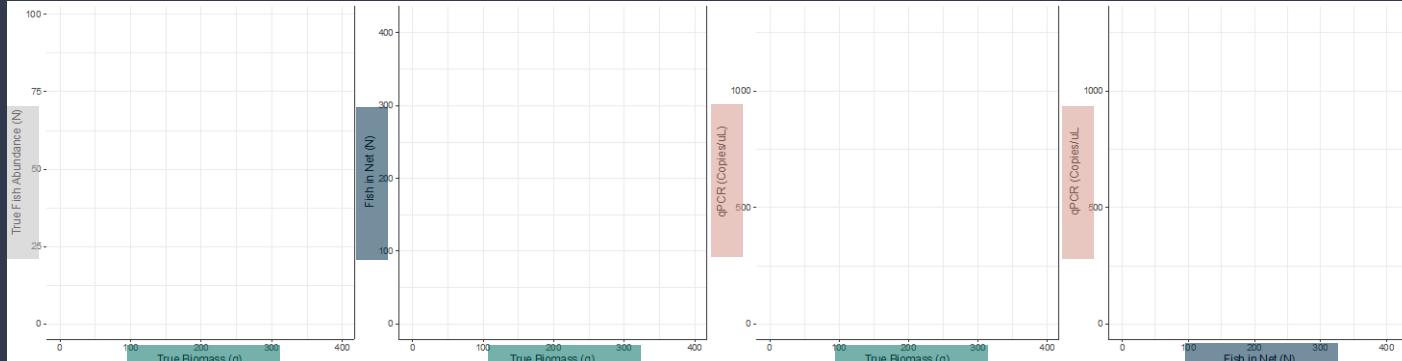
Reciprocal out-of-sample cross-validation

# Compounding Noise

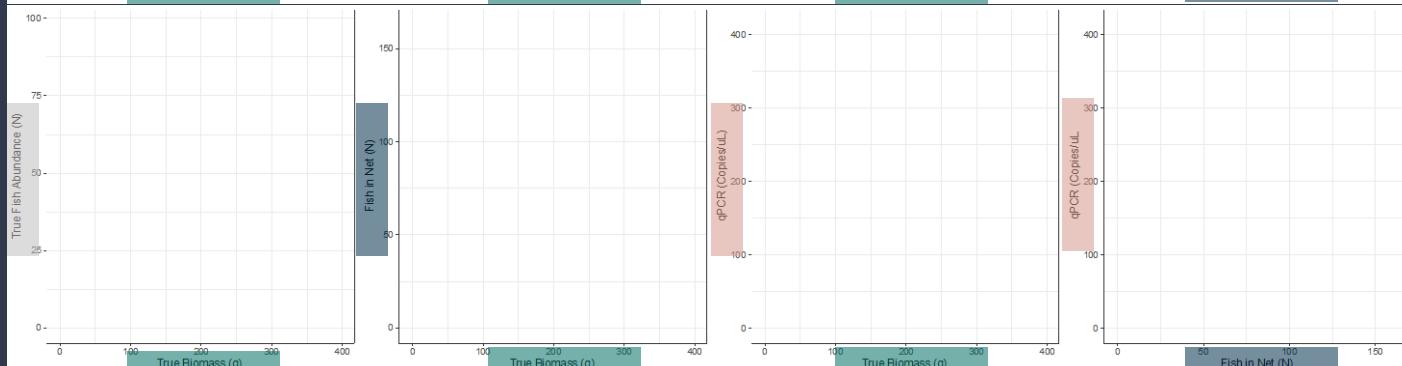
Variable mass-per-fish

1:1 eDNA copies per unit mass

Moderate  
Observation  
Variability



Low  
Observation  
Variability



The spatial/temporal scale of process + observation variability determine what correlations might exist